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ANSWER 1 OF 18 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN DUPLICATE 1

ACCESSION NUMBER: 2003:412775 BIOSIS DOCUMENT NUMBER: PREV200300412775

Nonlinear effects in subthreshold virtual electrode TITLE:

polarization.

Sambelashvili, Aleksandre T.; Nikolski, Vladimir P.; AUTHOR (S):

Efimov, Igor R. (1)

(1) Case Western Reserve Univ., 10900 Euclid Ave., CORPORATE SOURCE:

Wickenden Bldg., Rm. 520, Cleveland, OH, 44106-7207, USA:

ire@cwru.edu USA

American Journal of Physiology, (June 2003, 2003) Vol. 284, SOURCE:

No. 6 Part 2, pp. H2368-H2374. print.

ISSN: 0002-9513.

DOCUMENT TYPE: Article LANGUAGE: English

L2

We studied cardiac membrane polarization produced by subthreshold stimuli in 1) rabbit ventricular muscle using high-resolution fluorescent imaging with the voltage-sensitive dye

pyridinium 4-(2-(6-(dibutylamino)-2-naphthalenyl)-ethenyl)-1-(3sulfopropyl) hydroxide (di-4-ANEPPS) and 2) an active bidomain model with Luo-Rudy ion channel kinetics. Both in vitro and in

numero models show that the common dog-bone-shaped VEP is present at any stimulus strength during both systole and diastole. Diastolic subthreshold VEPs exhibited nonlinear properties that were expressed in

time-dependent asymmetric reversal of membrane polarization with respect to stimulus polarity. The bidomain model reveals that this asymmetry is due to nonlinear properties of the inward rectifier potassium current. Our results suggest that active ion channel kinetics

modulate the transmembrane polarization pattern that is predicted by the linear bidomain model of cardiac syncytium.

ACCESSION NUMBER: 2003:213163 BIOSIS DOCUMENT NUMBER: PREV200300213163

TITLE: Measurement of membrane potential from colonies of HEK293

cells transiently expressing ion channels by use of voltage-

sensitive fluorescent dye.

AUTHOR(S): Hotta, Aya (1); Ohya, Susumu (1); Muraki, Katsuhiko (1);

Imaizumi, Yuji (1)

CORPORATE SOURCE: (1) Dept. Mol. Cell. Pharmacol., Grad. Sch. Pharm. Sci.,

Nagoya City Univ., Nagoya, 467-8603, Japan Japan

SOURCE: Journal of Pharmacological Sciences, (2003) Vol. 91, No.

Supplement I, pp. 245P. print.

Meeting Info.: 76th Annual Meeting of the Japanese

Pharmacological Society Fukuoka, Japan March 24-26, 2003

Japanese Pharmacological Society

. ISSN: 1347-8613.

DOCUMENT TYPE: Conference LANGUAGE: English

TI Measurement of membrane potential from colonies of HEK293 cells

transiently expressing ion channels by use

of voltage-sensitive fluorescent dye.

L2 ANSWER 3 OF 18 CAPLUS COPYRIGHT 2003 ACS on STN

ACCESSION NUMBER: 2002:522155 CAPLUS

DOCUMENT NUMBER: 137:91389

TITLE: cDNAs encoding mammalian taste receptor cell- specific

ion channel subunits and screening for effectors of

taste signaling

INVENTOR(S): Zuker, Charles S.; Zhang, Yifeng

PATENT ASSIGNEE(S): The Regents of the University of California, USA

SOURCE: PCT Int. Appl., 306 pp.

CODEN: PIXXD2

DOCUMENT TYPE: Patent LANGUAGE: English

FAMILY ACC. NUM. COUNT: 1

PATENT INFORMATION:

PATENT NO.				KIND		DATE			APPLICATION NO.					DATE				
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REFERENCE COUNT: 5 THERE ARE 5 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

AB The invention includes nucleic acid and amino acid sequences of a mouse, human and rat taste cell-specific ion channel subunit that is specifically expressed in taste cells. Also provided are antibodies to such subunits, methods of detecting such nucleic acids and proteins, and methods of screening for modulators of taste cell specific ion channel subunit signaling. More specifically, taste cell-specific ion channels modulate the transmembrane Ca2+ ion flux which may be monitored by voltage clamp assays, patch clamp assays, radiolabeled ion flux assays or fluorescence assays using ion

## sensitive dyes.

ANSWER 4 OF 18 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN 1.2

DUPLICATE 2

2002:205100 BIOSIS ACCESSION NUMBER: PREV200200205100 DOCUMENT NUMBER:

TITLE:

SOURCE:

Transgenic mice expressing a pH and Cl- sensing

yellow-fluorescent protein under the control of a potassium

channel promoter.

Metzger, Friedrich; Repunte-Canonigo, Vez; Matsushita, AUTHOR (S):

Shinichi; Akemann, Walther; Diez-Garcia, Javier; Ho, Chi Shun; Iwasato, Takuji; Grandes, Pedro; Itohara, Shigeyoshi;

Joho, Rolf H.; Knopfel, Thomas (1)

CORPORATE SOURCE:

(1) Laboratory for Neuronal Circuit Dynamics, Brain Science

Institute, RIKEN, 2-1 Hirosawa, Wako-shi, Saitama,

351-0198: knopfel@brain.riken.go.jp Japan

European Journal of Neuroscience, (January, 2002) Vol. 15,

No. 1, pp. 40-50. http://www.blackwell-science.com/ cgilib/jnlpage.asp?Journal=ejn&File=ejn.print.

ISSN: 0953-816X.

DOCUMENT TYPE:

Article English

LANGUAGE: years a variety of genetically encodable optical probes that monitor AB. physiological parameters such as local pH, Ca2+, Cl-, or transmembrane voltage have been developed. These sensors are based on variants of green-fluorescent protein (GFP) and can be synthesized by mammalian . after transfection with cDNA. To use these sensor proteins in intact brain tissue, specific promoters are needed that drive protein expression at a sufficiently high expression level in distinct neuronal subpopulations. Here we investigated whether the promoter sequence of a particular potassium channel may be useful for this purpose. We produced transgenic mouse lines carrying the gene for enhanced yellow-fluorescent protein (EYFP), a yellow-green pHand Cl- sensitive variant of GFP, under control of the Kv3.1 K+ channel promoter (pKv3.1). Transgenic mouse lines displayed high levels of EYFP expression, identified by confocal microscopy, in adult cerebellar granule cells, interneurons of the cerebral cortex, and in neurons of hippocampus and thalamus. Furthermore, using living cerebellar slices we demonstrate that expression levels of EYFP are sufficient to report intracellular pH and Cl- concentration using imaging techniques and conditions analogous to those used with conventional ion-sensitive dyes. We conclude that transgenic mice expressing GFP-derived sensors under the control of cell-type specific promoters, provide a unique opportunity for functional characterization of defined subsets of.

ANSWER 5 OF 18 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER:

2001:540671 BIOSIS

DOCUMENT NUMBER:

PREV200100540671

TITLE:

Design and characterization of a DNA encoded voltage

sensitive fluorescent protein.

AUTHOR (S):

Knopfel, T. (1); Repunte-Canonigo, V. (1); Raj, C. D. (1);

Sakai, R. (1)

CORPORATE SOURCE:

(1) Laboratory for Neuronal Circuit Dynamics, Brain Science

Institute, RIKEN, Wako-shi Japan

SOURCE:

Society for Neuroscience Abstracts, (2001) Vol. 27, No. 2,

pp. 1583. print.

Meeting Info.: 31st Annual Meeting of the Society for Neuroscience San Diego, California, USA November 10-15,

2001

ISSN: 0190-5295.

DOCUMENT TYPE:

Conference

LANGUAGE:

English

SUMMARY LANGUAGE:

English

. . suggested as a promising approach to investigate the multineuronal AB. representation of information processing in brain tissue. However, intrinsic or extrinsic dye-mediated optical signals are often of limited use due to their slow response dynamics, low effective sensitivity, toxicity or undefined cellular origin. Protein-based and DNA encoded voltage sensors could overcome these limitations. Here we report the design and generation of a voltage sensitive fluorescent protein (VSFP1) consisting of a voltage sensing domain of a potassium channel and a pair of cyan and yellow mutants of green fluorescent protein (GFP). Modulation of fluorescence intensity by membrane potential was investigated in voltage-clamped HEK cells expressing VSFP1. Depolarizing voltage jumps resulted in an increase in the emission by YFP (>530 nm) with excitation of CFP (432 nm) while hyperpolarization of the membrane resulted in a decrease in fluorescence output. The current-tovoltage relationship of HEK cells expressing VSFP1 did not differ from that of untransfected cells demonstrating that VSFP1 did not form functional ion-conducting channels. The relationship between voltage change and fluorescence change was close to linear (r=0.99) with a slope of 1.8+-0.1%/100 mV (n=11 cells). In parallel measurements using the prototypic conventional voltage sensitive dye di-4-ANEPPS, we obtained a sensitivity of -5.3+-0.3%/100 mV from clean HEK cell membranes. The optical signals responded in the millisecond time scale of fast electrical signaling and are large enough to allow monitoring voltage changes at the single cell level.

DUPLICATE 3 ANSWER 6 OF 18 MEDLINE on STN

ACCESSION NUMBER:

2001164445 MEDITNE

21163415 PubMed ID: 11265727

DOCUMENT NUMBER: TITLE:

Cellular basis for dispersion of repolarization underlying

reentrant arrhythmias.

Akar F G; Laurita K R; Rosenbaum D S

CORPORATE SOURCE:

Department of Medicine, Heart and Vascular Research Center,

Case Western Reserve University, Cleveland, OH 44109-1998,

SOURCE:

JOURNAL OF ELECTROCARDIOLOGY, (2000) 33 Suppl 23-31. Ref:

Journal code: 0153605. ISSN: 0022-0736.

PUB. COUNTRY:

United States

DOCUMENT TYPE:

Journal; Article; (JOURNAL ARTICLE)

General Review; (REVIEW)

(REVIEW, TUTORIAL)

LANGUAGE:

English

FILE SEGMENT:

Priority Journals

ENTRY MONTH:

200104

ENTRY DATE:

Entered STN: 20010502

Last Updated on STN: 20010502 Entered Medline: 20010426

AB Substantial heterogeneity in ion channel density and expression exists in cells isolated from various regions of the heart. Cell-to-cell coupling in the intact heart, however, is expected to attenuate the functional expression of the ion channel heterogeneities. Due to limitations of conventional electrophysiological recording techniques, the extent to which cellular electrical heterogeneities are functionally present in intact myocardium remains unknown. High-resolution optical mapping with voltagesensitive dyes was used to measure transepicardial and transmural repolarization gradients in the Langendorff perfused guinea pig ventricle and the canine wedge.

ANSWER 7 OF 18 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN ACCESSION NUMBER:

2001:88917 BIOSIS

DOCUMENT NUMBER:

PREV200100088917

TITLE:

Mechanisms of hypoxic excitation of vasomotor neurons of

rostral ventrolateral medulla.

AUTHOR (S):

Wang, G. (1); Zhou, P.; Repucci, M.; Reis, D. J.

CORPORATE SOURCE: SOURCE:

(1) Weill Med. Coll. of Cornell Univ., New York, NY USA Society for Neuroscience Abstracts, (2000) Vol. 26, No.

1-2, pp. Abstract No.-443.11. print.

Meeting Info.: 30th Annual Meeting of the Society of Neuroscience New Orleans, LA, USA November 04-09, 2000

Society for Neuroscience

. ISSN: 0190-5295.

DOCUMENT TYPE:

Conference English

LANGUAGE: English SUMMARY LANGUAGE: English

AB. . . excited by hypoxia, initiating patterned autonomic responses of O2-conserving (diving) reflex. The O2-sensing in peripheral chemoreceptor cells is associated with O2-sensitive K+ channel

activity. We investigated whether hypoxic excitation of RVLM neurons results from activation or inhibition of O2-sensitive,

voltage-gated ion channels. RVLM neurons from

3-11 day old rat pups were retrogradely labeled with rhodamine-labeled dyes injected into the T2-T4 spinal segment. Brainstem slices (150-200mu) were obtained and RVLM neurons identified under

epifluorescence. The labeled RVLM. . . 2.2 mV (n=3, p<0.05) without any SD by 125 muM NaCN. To determine if this hypoxic effect is related to O2-sensitive voltage-gated ion channels

, the ion currents of RVLM neurons were recorded using the whole-cell voltage-clamp. While the Na+, A-type K+ and Ca++ currents were not significantly affected by NaCN, a sustained outward K+ current was. . . controls to 947 +- 109.8 pA of NaCN-treated neurons (n=4, p<0.01). Post-recording single cell RT-PCR was also conducted. RVLM neurons expressed TH and O2-sensitive, voltage -gated K+ channels Kv2.1 and Kv 3.1. These results suggest that

inhibition of O2-sensitive K+ channels might contribute to hypoxic excitation of RVLM neurons.

L2 ANSWER 8 OF 18 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN DUPLICATE 4

ACCESSION NUMBER: 1999:37
DOCUMENT NUMBER: PREV199

1999:374940 BIOSIS PREV199900374940

TITLE:

Modulation of glioma cell migration and invasion using Cl-

and K+ ion channel blockers.

AUTHOR (S):

Soroceanu, Liliana; Manning, Timothy J., Jr.; Sontheimer,

Harald (1)

CORPORATE SOURCE:

(1) 1719 6th Avenue South CIRC 545, Birmingham, AL,

35294-0021 USA

SOURCE:

Journal of Neuroscience, (July 15, 1999) Vol. 19, No. 14,

pp. 5942-5954. ISSN: 0270-6474.

DOCUMENT TYPE: Article
LANGUAGE: English
SUMMARY LANGUAGE: English

AB. . . Mechanisms that allow glioma cells to disseminate, migrating through the narrow extracellular brain spaces are poorly understood. We recently demonstrated expression of large voltage-dependent chloride (Cl-) currents, selectively expressed by human glioma cells in vitro and in situ (Ullrich et al., 1998). Currents are sensitive to several Cl- channel blockers, including chlorotoxin (Ctx), (Ullrich and Sontheimer, 1996; Ullrich et al., 1996), tetraethylammonium chloride (TEA), and tamoxifen (Ransom and Sontheimer, 1998). Using Transwell migration assays, we show that blockade of glioma Cl- channels specifically inhibits tumor cell migration in a dose-dependent manner. Ctx (5 muM), tamoxifen (10 muM), and TEA (1 mM) also. . . brain aggregates, used as an in vitro model to assess tumor invasiveness. Anion replacement studies suggest that permeation of

chloride ions through glioma chloride channel is obligatory for cell migration. Osmotically induced cell swelling and subsequent regulatory volume decrease (RVD) in cultured glioma cells were. in glioma cells were inhibited by 5 muM Ctx, 10 muM tamoxifen, and 1 mM TEA, as determined using the Cl-sensitive fluorescent dye 6-methoxy-N-ethylquinolinium iodide. Collectively, these data suggest that chloride channels in glioma cells may enable tumor invasiveness, presumably by facilitating cell shape and cell volume changes that are more conducive.

ANSWER 9 OF 18 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

DUPLICATE 5

ACCESSION NUMBER: 1999:351188 BIOSIS PREV199900351188 DOCUMENT NUMBER:

Block by ruthenium red of cloned neuronal voltage-gated TITLE:

calcium channels.

Cibulsky, Susan M.; Sather, William A. (1) AUTHOR(S):

(1) Neuroscience Center, B-138, University of Colorado Health Sciences Center, 4200 E. 9th Ave., Denver, CO, 80262 CORPORATE SOURCE:

Journal of Pharmacology and Experimental Therapeutics, SOURCE:

(June, 1999) Vol. 289, No. 3, pp. 1447-1453.

ISSN: 0022-3565.

DOCUMENT TYPE: Article LANGUAGE: SUMMARY LANGUAGE:

English English

The dye ruthenium red (RuR) has diverse experimental uses, including block of ion channels. RuR is a well described antagonist of one class of intracellular Ca2+ release channels, the ryanodine receptors, but recently this compound has also been identified as a putative blocker of voltage-gated calcium channels of the surface membrane involved in neurotransmitter release. Using electrophysiological methods, we have studied the action of RuR upon pure populations of neuronal voltage-gated ion channels heterologously expressed in Xenopus laevis oocytes. All four channel types studied, including class A (P/Q-type), class B (N-type), class C (L-type), and class E channels, are sensitive to RuR, with IC50 values ranging from 0.7 to 67.1 muM. Block of class C and class E channels most likely results from 1:1 binding of ruthenium red at a site in the extracellular entrance to the pore, resulting in obstruction of permeant ion flux through these channels

. The mechanism of block of classA and class B channels is more complex, requiring binding of more than one molecule of RuR per channel.

ANSWER 10 OF 18 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN **DUPLICATE 6** 

ACCESSION NUMBER: DOCUMENT NUMBER:

1998:483420 BIOSIS PREV199800483420

TITLE:

Rapid Ca2+ entry through Ca2+-permeable AMPA/kainate channels triggers marked intracellular Ca2+ rises and

consequent oxygen radical production.

AUTHOR (S):

Carriedo, Sean G.; Yin, Hong Zhen; Sensi, Stefano L.;

Weiss, John H. (1)

CORPORATE SOURCE:

(1) Dep. Neurology, Univ. California, Irvine, Irvine, CA

92697-4292 USA

SOURCE:

Journal of Neuroscience, (Oct. 1, 1998) Vol. 18, No. 19,

pp. 7727-7738. ISSN: 0270-6474.

DOCUMENT TYPE:

Article

LANGUAGE: English

The widespread neuronal injury that results after brief activation of highly Ca2+-permeable NMDA channels may, in large part, reflect

mitochondrial Ca2+ overload and the consequent production of injurious oxygen radicals. In contrast, AMPA/kainate receptor. . . studies have not found evidence of comparable oxygen radical production. Subsets of central neurons, composed mainly of GABAergic inhibitory interneurons, express AMPA/kainate channels that are directly permeable to Ca2+ ions. Microfluorometric techniques were performed by using the oxidation-sensitive dye hydroethidine (HEt) to determine whether the relatively rapid Ca2+ flux through AMPA/kainate channels expressed on GABAergic neurons results in oxygen radical production comparable to that triggered by NM DA. Consistent with previous studies, NMDA. . . triggered increases in fluorescence in most cultured cortical neurons, whereas high K+ (50 mM) exposures (causing depolarization-induced Ca2+ influx through voltage-sensitive Ca2+ channels) caused little fluorescence change. In contrast, kainate exposure caused fluorescence increases in a distinct subpopulation of neurons; immunostaining for glutamate. . . oxygen radical production paralleled the effect of these exposures on intracellular Ca2+ levels when they were monitored with the low-affinity Ca2+-sensitive dye fura-2FF, but not with the high-affinity dye fura-2. Inhibition of mitochondrial electron transport with CN- or rotenone almost completely blocked kainate-triggered oxygen radical production. Furthermore, antioxidants attenuated. . . resulting from brief exposures of NMDA or kainate. Thus, as with NMDA receptor activation, rapid Ca2+ influx through Ca2+-permeable AMPA/kainate channels also may result in mitochondrial Ca2+ overload and consequent injurious oxygen radical production.

L2 ANSWER 11 OF 18 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: 1998:484015 BIOSIS DOCUMENT NUMBER: PREV199800484015

TITLE: Voltage-activated currents in identified giant interneurons

isolated from adult crickets Gryllus bimaculatus.

AUTHOR(S): Kloppenburg, Peter; Hoerner, Michael (1)

CORPORATE SOURCE: (1) Inst. Zool. Anthropol., Dep. Cell Biol., Univ.

Goettingen, Berliner Strasse 28, D-37073 Goettingen Germany

SOURCE: Journal of Experimental Biology, (Sept., 1998) Vol. 201,

No. 17, pp. 2529-2541.

ISSN: 0022-0949.

DOCUMENT TYPE: Article LANGUAGE: English

bodies was established. Prior to cell dissociation, the giant interneurons were backfilled through their axons in situ with a fluorescent dye (dextran tetramethylrhodamine). In primary cell cultures, the cell bodies of giant interneurons were identified among a population of co-cultured neurons. . . their red fluorescence. Action potentials were recorded from the cell bodies of the cultured interneurons suggesting that several types of voltage-activated ion channels exist in these cells. Using voltage-clamp recording techniques, four voltage-activated currents were isolated and characterized. The giant interneurons express at least two distinct K+ currents: a transient current that is blocked by 4-aminopyridine (4 X 10-3 mol-1) and a. . partially blocked by tetraethylammonium (3 X 10-2 mol-1) and quinidine (2 X 10-4 mol-1). In addition, a transient Na+ current sensitive to 10-7 mol 1-1 tetrodotoxin and a Ca2+ current blocked by 5 X 10-4 mol 1-1 CdCl2 have been characterized...

L2 ANSWER 12 OF 18 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN DUPLICATE 7

ACCESSION NUMBER: 1996:37100 BIOSIS DOCUMENT NUMBER: PREV199698609235

TITLE: Activation of nicotinic acetylcholines receptors expressed in quail fibroblasts: Effects on intracellular calcium.

AUTHOR(S): Cross, K. M. L. (1); Jane, S. D.; Wild, A. E.; Foreman, R.

C.; Chad, J. E.

CORPORATE SOURCE: (1) Dep. Physiol. Pharmacol., University Southampton,

Southampton SO16 7PX UK

SOURCE: British Journal of Pharmacology, (1995) Vol. 116, No. 7,

pp. 2838-2844. ISSN: 0007-1188.

DOCUMENT TYPE: Article LANGUAGE: English

AB 1 The aim of these experiments was to determine the ability of the

muscle-type nicotinic acetylcholine receptor (AChR) stably expressed in quail fibroblasts (QF18 cells) to elevate

intracellular calcium ((Ca-2+)-i) upon activation. Ratiometric confocal microscopy, with the calcium-sensitive fluorescent dye

Indo-1 was used. 2 Application of the nicotinic agonist, suberyldicholine (SDC), to the transfected QF18 cells caused an increase in. . . were

blocked by prior application of alpha-bungarotoxin (200 nM), by the

addition of Ca-2+ (100 mu-M), by removal of Na+ ions from the

extracellular solution, or by the voltage-sensitive

calcium channel blockers nifedipine and omega-conotoxin, which act with IC-50 values of 100 nM and 100 pM respectively. 5 We conclude

that activation of the nicotinic AChRs leads to a Na+-dependent depolarization and hence activation of endogenous voltage-

sensitive Ca-2+ channels in the plasma membrane and an increase in (Ca-2+)-i. There is no significant entry of Ca-2+ through the

L2 ANSWER 13 OF 18 MEDLINE On STN DUPLICATE 8

ACCESSION NUMBER: 96342976 MEDLINE

DOCUMENT NUMBER: 96342976 PubMed ID: 8741761

TITLE: Zn(2+) permeates Ca(2+) permeable AMPA/kainate channels and

triggers selective neural injury.

AUTHOR: Yin H Z; Weiss J H

CORPORATE SOURCE: Department of Neurology, University of California, Irvine,

92717-4290, USA.

CONTRACT NUMBER: AG00495 (NIA)

nicotinic receptor.

NS30884 (NINDS)

SOURCE: NEUROREPORT, (1995 Dec 15) 6 (18) 2553-6.

Journal code: 9100935. ISSN: 0959-4965.

PUB. COUNTRY: ENGLAND: United Kingdom

DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals

ENTRY MONTH: 199610

ENTRY DATE: Entered STN: 19961106

Last Updated on STN: 19970203 Entered Medline: 19961024

AB . . . Brief exposures of cortical cultures to kainate (100 mu M) plus Zn(2+) (300 mu M) cause fluorescence of the Zn(2+) sensitive dye, TS-Q, to appear in virtually all neurons, probably reflecting depolarization and secondary Zn(2+) permeation through voltagesensitive Ca(2+) channels. However, if Na+ ions are removed from the media (to prevent depolarization), prominent TS-Q fluorescence is still observed in the small subset of neurons labeled by kainate stimulated Co(2+) uptake (Co(2+)(+) neurons), a histochemical technique that identifies neurons expressing Ca(2+) permeable AMPA/kainate receptor-gated channels. Kainate/Zn(2+) exposures in Na+ containing media with lower (50-100 mu M) Zn(2+) concentrations resulted 24 h later in selective loss. .

L2 ANSWER 14 OF 18 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN DUPLICATE 9

ACCESSION NUMBER: 1994:129924 BIOSIS DOCUMENT NUMBER: PREV199497142924

Cadmium Toxicity in Rat Pheochromocytoma Cells: Studies on TITLE:

the Mechanism of Uptake.

Hinkle, Patricia M. (1); Osborne, Matthew E. AUTHOR (S):

(1) Dep. Pharmacology, University Rochester School Medicine CORPORATE SOURCE:

and Dentistry, Rochester, NY 14642 USA

Toxicology and Applied Pharmacology, (1994) Vol. 124, No. SOURCE:

1, pp. 91-98. ISSN: 0041-008X.

DOCUMENT TYPE: Article English LANGUAGE:

The uptake and toxicity of cadmium were compared in two rat AB

pheochromocytoma cell lines: PC12 cells, which express

voltage-sensitive calcium channels, and PC18 cells, which do not. PC12 but not PC18 cells responded to depolarization with an increase in 45Ca-2+ uptake and an increase in the concentration of cytoplasmic free calcium ion, (Ca-2+). These responses were blocked by the dihydropyridine calcium channel antagonist nimodipine and amplified by the agonist BAY K8644, drugs selective for L-type channels. Cadmium caused death of PC12 cells with an LC50 of 12 mu-M. Inclusion of high K+ with the agonist BAY. . . = 6 mu-M), whereas nimodipine protected against cadmium toxicity (LC50 = 30 mu-M). In contrast, drugs acting on L-type calcium channels did not affect Cd-2+ toxicity for PC18 cells (LC50 15 mu-M). Fura 2 was used to measure intracellular free Cd-2+. . . PC12 cells. Cd-2+ fluorescence appeared to be concentrated near the plasma membrane. The results confirm the potential involvement of calcium channels in cadmium transport and extend the use of intracellularly trapped fluorescent dyes to monitor intracellular free cadmium ion concentration.

ANSWER 15 OF 18 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

DUPLICATE 10

ACCESSION NUMBER: 1993:272255 BIOSIS DOCUMENT NUMBER: PREV199396002480

Neural induction suppresses early expression of the TITLE:

inward-rectifier potassium channel in the ascidian

blastomere.

AUTHOR(S): Okamura, Yasushi (1); Takahashi, Kunitaro

(1) Dep. Neurobiol., Inst. Brain Res., Fac. Med., Univ. Tokyo, Hongo 7-3-1, Bunkyo-ku, Tokyo Japan CORPORATE SOURCE:

Journal of Physiology (Cambridge), (1993) Vol. 463, No. 0, SOURCE:

pp. 245-268.

ISSN: 0022-3751.

DOCUMENT TYPE: Article LANGUAGE: English

1. Early expression of ion channels

following neural induction was examined in isolated, cleavage-arrested blastomeres from the ascidian embryo using a two-electrode voltage clamp. Currents were recorded from the isolated, cleavage-arrested blastomere, a-4-2, after treatment with serine protease, subtilisin, which induces neural differentiation as consistently as cell contact. 2. The inward-rectifier K+ current increased at the late gastrula stage shortly after the sensitive period for neural inducation both in the induced (protease-treated) and uninduced cells. Ca-2+ channels, characteristic of epidermal-type differentiation, and delayed-rectifier K+ channels and differentiated-type Na+ channels, characteristic of neural-type differentiation appeared much later than the inward-rectifier K+ channels, at a time corresponding to the tail bud stage of the intact embryo. 3. When cells were treated with subtilisin. . = 14) than in untreated cells (11.25 + - 3.10 nA, n =26). The same changes in the inward-rectifier K+ channel were also observed in a-4-2 blastomeres which were induced by cell contact with an A-4-1 blastomere. However, when cells were. . . period for neural induction, the amplitude of the inward-rectifier K+ current was the same as in untreated cells. Thus the expressed level of the

inward-rectifier K+ channel was linked to the determination of neural or epidermal cell types. 4. There was no significant difference in the input. . . uninduced cells, indicating that the difference in the amplitude of the inward-rectifier K+ currents derived from a difference in the channel density rather than a difference in cell surface area. 5. The expression of the inward-rectifier K+ channel at the late gastrula stage was sensitive to alpha-amanitin, a highly specific transcription inhibitor. In both induced and uninduced cells, injection of alpha-amanitin at the 32-cell stage reduced the current density of the inward-rectifier K+ channel to about 2 nA/nF, corresponding to 13% of that recorded from uninjected cells. By contrast, the expression of the fast-inactivating-type Na+ current, which transiently increased along with the inward-rectifier K+ channel, was resistant to alpha-amanitin injection. 6. The dose of alpha-amanitin injected was controlled by monitoring co-injected fluorescent dye, fura-2. The dose of alpha-amanitin required for 50% suppression of the inward-rectifier K+ current was 3.0 ng/ml. This was close. . . was taken into account. 7. In the uninduced cells, injection of alpha-amanitin later than the 32-cell stage partially suppressed the expression of the inward-rectifier K+ channel and the fraction of suppression was related linearly to the time of injection. By contrast, in protease-treated cells (induced cells) the expression of the inward-rectifier K+ channel depended only on transcription before protease treatment. We concluded that inductive signals suppressed transcription of the inward-rectifier K+ channel which had already started before the 64-cell stage.

L2 ANSWER 16 OF 18 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: 1988:175601 BIOSIS

DOCUMENT NUMBER: BA85:87703

TITLE: FLOW CYTOMETRIC ANALYSIS OF MEMBRANE POTENTIAL IN EMBRYONIC

RAT SPINAL CORD CELLS.

AUTHOR(S): MANDLER R N; SCHAFFNER A E; NOVOTNY E A; LANGE G D; BARKER

JL

CORPORATE SOURCE: LAB. NEUROPHYSIOL., NATL. INST. NEUROL. COMMUN. DISORDERS

STROKE, BUILD. 36, ROOM 2CO2, NIH, BETHESDA, MD. 20892.

SOURCE: J NEUROSCI METHODS, (1988) 22 (3), 203-214.

CODEN: JNMEDT. ISSN: 0165-0270.

FILE SEGMENT: BA; OLD LANGUAGE: English

. . potential in suspensions of embryonic rat spinal cord cells was carried out in a fluorescence-activated cell sorter (FACS) using anionic voltage-sensitive, fluorescent dyes (oxonols). The FACS or flow cytometer is an analytical instrument that measures optical properties of large cell populations at a. . . is directly related to the degree of cell depolarization. Incubation of cells in elevated K+ concentrations or with the Na+ channel agonist batrachotoxin (BTX) changed the fluorescence intensity distribution pattern of the live-cell population; these changes were consistent with the depolarizing. . . in the dead-cell population. The BTX-induced shift was blocked by tetrodotoxin (TTX) and was reversed in Na+-free medium, indicating embryonic expression of functional Na+ channels. Fluorescence microscopy of sorted cells showed that live cells typically exhibited circumferential ring-like patterns, whose intensities were enhanced under depolarizing conditions. The results show that flow cytometry combined with oxonol dyes can be used to measure the relative membrane potential of large numbers of individual central nervous system cells. The analysis of the changes in the distributions of these membrane potentials can be used to reveal the development of functional ion conductance mechanisms.

L2 ANSWER 17 OF 18 EMBASE COPYRIGHT 2003 ELSEVIER INC. ALL RIGHTS RESERVED. on STN

ACCESSION NUMBER: 88006778 EMBASE

DOCUMENT NUMBER: 1988006778

Flow cytometric analysis of membrane potential in embryonic TITLE:

rat spinal cord cells.

AUTHOR: Mandler R.N.; Schaffner A.E.; Novotny E.A.; Lange G.D.;

Barker J.L.

Laboratory for Neurophysiology, National Institute of CORPORATE SOURCE:

Neurological and Communicative Disorders and Stroke,

National Institutes of Health, Bethesda, MD 20892, United

States

Journal of Neuroscience Methods, (1987) 22/3 (203-213). SOURCE:

ISSN: 0165-0270 CODEN: JNMEDT

COUNTRY: Netherlands

DOCUMENT TYPE: Journal

FILE SEGMENT: 002 Physiology

052 Toxicology

LANGUAGE: English SUMMARY LANGUAGE: English

. . potential in suspension of embryonic rat spinal cord cells was carried out in a fluorescence-activated cell sorter (FACS) using anionic voltage-sensitive, fluorescent dyes (oxonols).

The FACS or flow cytometer is an analytical instrument that measures optical properties of large cell populations at a. . . is directly related to the degree of cell depolarization. Incubation of cells in elevated K+ concentrations or with the Na+ channel agonist

batrachotoxin (BTX) changed the fluorescence intensity distribution pattern of the live-cell population; these changes were consistent with the depolarizing. . . in the dead-cell population. The BTX-induced shift was blocked by tetrodotoxin (TTX) and was reversed in Na+-free medium, indicating embryonic expression of functional Na+

channels. Fluorescence microscopy of sorted cells showed that live cells typically exhibited circumferential ring-like patterns, whose intensities were enhanced under depolarizing conditions. The results show that flow cytometry combined with oxonol dyes can be used to measure the relative membrane potential of large numbers of individual central nervous system cells. The analysis of the changes in the distributions of these membrane potentials can be used to reveal the development of functional ion conductance mechanisms.

ANSWER 18 OF 18 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN **DUPLICATE 11** 

ACCESSION NUMBER: 1981:183027 BIOSIS

DOCUMENT NUMBER: BA71:53019

TITLE: INTRA CELLULAR CALCIUM ACCUMULATION DURING DE POLARIZATION

IN A MOLLUSCAN NEURON. GORMAN A L F; THOMAS M V

CORPORATE SOURCE: DEPARTMENT OF PHYSIOLOGY, BOSTON UNIVERSITY SCHOOL OF

> MEDICINE, BOSTON, MASSACHUSETTS 02118, USA. J PHYSIOL (LOND), (1980) 308 (0), 259-286.

CODEN: JPHYA7. ISSN: 0022-3751.

FILE SEGMENT: BA; OLD LANGUAGE: English

AUTHOR (S):

SOURCE:

The bursting pacemaker neuron R-15 of Aplysia [A. californica] was

injected with the Ca2+ sensitive dye arsenazo III.

Changes in absorbance were measured with a differential spectrophotometer to monitor changes in free intracellular Ca2+ during membrane

depolarization under voltage clamp conditions. Dye

absorbance increased linearly for depolarizing pulse durations up to 100 ms and approximately linearly between 100-300 ms, but for longer durations the absorbance change decreased. The absorbance change vs. voltage relation increased steeply between -20 and 0 mV (e-fold/8.5 mV), peaked at +36 mV and declined nonlinearly to an estimated. . . null or

suppression potential of about +139 mV. Tetrodotoxin (5 .times. 10-5 M) had no effect on the change in dye absorbance produced by brief

or long duration stimuli whereas Ca2+ free [artificial seawater] abolished

all changes in dye absorbance. The absorbance change saturated with increasing external Ca2+ concentrations. The relation between dye absorbance and external Ca2+ concentration was hyperbolic and for a small range of external Ca2+ concentration and membrane potentials could be fitted by a Michaelis-Menten expression where the dissociation constant and the maximum absorbance change are voltage dependent. The absorbance change was reduced by external divalent ions which block the Ca2+ channel (e.g., Cd2+ and Ni2+). The suppression of dye absorbance was increased by membrane depolarization and suggests that there is a voltage dependent site within the Ca2+ channel which binds divalent ions. The decline of the absorbance-voltage relation from its peak to the suppression potential showed a greater nonlinearity when longer duration voltage clamp pulses were used. The nonlinearity can be explained if the accumulation of Ca2+ ions next to the inner surface of the membrane during depolarization reduces the driving force on Ca2+ ions decreasing Ca2+ ion influx. The suppression potential estimated from the absorbancevoltage relation increased 29 mV/10-fold change in the external Ca2+ concentration and can be used to estimate the Ca2+ equilibrium potential. The change in dye absorbance produced by brief depolarizing voltage clamp steps was inactivated at positive holding potentials (50% inactivation at about -14 mV). The slow decrease in dye absorbance during prolonged depolarization probably is caused by inactivation of the Ca2+ channel.

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=> s (voltage (s) sensitive (s) dye) (p) ( recombina? (s) ion (s) channel)
             1 (VOLTAGE (S) SENSITIVE (S) DYE) (P) (RECOMBINA? (S) ION (S)
               CHANNEL)
```

## => d 13 total ibib kwic

ANSWER 1 OF 1 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER:

2001:417367 BIOSIS DOCUMENT NUMBER: PREV200100417367

TITLE:

Usefulness and limitation of DiBAC4(3), a voltage-sensitive fluorescent dye, for the measurement of membrane potentials regulated by recombinant large conductance Ca2+-activated

K+ channels in HEK293 cells.

AUTHOR (S): Yamada, Aki; Gaja, Norikazu; Ohya, Susumu; Muraki,

Katsuhiko; Narita, Hiroshi; Ohwada, Tomohiko; Imaizumi,

Yuji (1)

CORPORATE SOURCE:

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Nagoya City University, Nagoya, 467-8603:

yimaizumi@phar.nagoya-cu.ac.jp Japan

SOURCE:

Japanese Journal of Pharmacology, (July, 2001) Vol. 86, No.

3, pp. 342-350. print.

ISSN: 0021-5198.

DOCUMENT TYPE: LANGUAGE:

Article English

SUMMARY LANGUAGE: English

Major Concepts Biochemistry and Molecular Biophysics; Pharmacology

IT Chemicals & Biochemicals

DiBAC4(3) [bis-(1,3-dibutylbarbituric acid)-trimethine oxonol]:

limitation, usefulness, voltage-sensitive

fluorescent dye; Evans blue: BK channel opener; NS-1619;

recombinant large conductance calcium(II)-activated potassium ion channels